Devilishly radical NETwork in COVID-19: Oxidative stress, neutrophil extracellular traps (NETs), and T cell suppression

Günther Schönrich a, **, Martin J. Raftery a, Yvonne Samstag b, *

a Institute of Virology, Charité – Universitätsmedizin Berlin, Free Universität Berlin, Humboldts-Universität zu Berlin, Berlin Institute of Health, Berlin, Germany
b Section Molecular Immunology, Institute of Immunology, Heidelberg University Hospital, Heidelberg, Germany

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ABSTRACT
Pandemic coronavirus disease 2019 (COVID-19) is caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) and poses an unprecedented challenge to healthcare systems due to the lack of a vaccine and specific treatment options. Accordingly, there is an urgent need to understand precisely the pathogenic mechanisms underlying this multifaceted disease. There is increasing evidence that the immune system reacts insufficiently to SARS-CoV-2 and thus contributes to organ damage and to lethality. In this review, we suggest that the overwhelming production of reactive oxygen species (ROS) resulting in oxidative stress is a major cause of local or systemic tissue damage that leads to severe COVID-19. It increases the formation of neutrophil extracellular traps (NETs) and suppresses the adaptive arm of the immune system, i.e. T cells that are necessary to kill virus-infected cells. This creates a vicious cycle that prevents a specific immune response against SARS-CoV-2. The key role of oxidative stress in the pathogenesis of severe COVID-19 implies that therapeutic counterbalancing of ROS by antioxidants such as vitamin C or NAC and/or by antagonizing ROS production by cells of the mononuclear phagocyte system (MPS) and neutrophil granulocytes and/or by blocking of TNF-α can prevent COVID-19 from becoming severe. Controlled clinical trials and preclinical models of COVID-19 are needed to evaluate this hypothesis.

1. Introduction

The present pandemic represents the third known outbreak of a coronavirus (CoV)-associated severe human disease due to spillover of an animal coronavirus to humans (Wu et al., 2020). The first time this group of viruses entered the public spotlight was in 2002 during the Severe Acute Respiratory Syndrome (SARS) epidemic caused by SARS-CoV (Drosten et al., 2003). Subsequently, in 2012 the Middle East respiratory syndrome (MERS)-CoV was identified in patients (Zaki et al., 2012). Finally, SARS-CoV-2 was discovered and associated with severe coronavirus disease 2019 (COVID-19) (Gorbalenya et al., 2020; Wu et al., 2020; Zhou et al., 2020b).

SARS-CoV, MERS-CoV, and SARS-CoV-2 belong to a family of enveloped positive-strand RNA viruses which infect vertebrates. There are also four endemic coronaviruses that are known to cause the common cold, a mild human disease of the upper-respiratory tract (Corman et al., 2018; Fung and Liu, 2019). SARS-CoV and SARS-CoV-2 are closely related with 79% genome sequence identity.

* Corresponding author.
** Corresponding author.
E-mail addresses: guenther.schoenrich@charite.de (G. Schönrich), yvonne.samstag@urz.uni-heidelberg.de (Y. Samstag).
3. Clinical course of COVID-19

The clinical course of SARS-CoV-2 infection is highly variable and ranges from asymptomatic or oligosymptomatic to severe organ dysfunction and death. In the incubation period and the early phase of COVID-19, SARS-CoV-2 replication in the upper respiratory tract triggers only a limited innate response. At this point, symptoms may be absent and viral spread may be terminated. The exact proportion of subclinical infections has to be determined in future serological studies. In the first week, however, non-productive cough, achings limbs, general fatigue, sore throat, loss of smell/taste, and headache may occur. Subsequently, SARS-CoV-2 can reach the lower respiratory tract and the lung alveoli (Li et al., 2020d). Whereas severe IAV infection is usually observed within a few days, severe COVID-19 occurs in the second symptomatic week and patients are often hospitalized for 3–4 days before being transferred to the intensive care unit (ICU). A characteristic feature found in chest computed tomography (CT) of acute lung injury (ALI) in severe COVID-19 are bilateral patchy infiltrates and “ground glass” opacities (Ye et al., 2020). Shortness of breath and fever may be a harbinger of severe COVID-19, which is complicated in 15%–30% of cases by an acute respiratory distress syndrome (ARDS) (Chen et al., 2020b; Guan et al., 2020; Huang et al., 2020; Wang et al., 2020a; Zhou et al., 2020a). A plethora of studies have shown that SARS-CoV-2 is not restricted to lung tissue but spreads systemically thereby causing an extensive immune response and considerable damage in other organs including brain, heart and blood vessels, liver, kidneys, and intestine (Li et al., 2020d; Shi et al., 2020). ARDS requires admission to the ICU, and is often accompanied by microthrombosis resulting in shock and multiorgan failure. In fact, acute cardiovascular and kidney damage are frequently observed in patients with severe COVID-19 and contribute to the case fatality rate (Akhermerov and Marban, 2020; Bonov et al., 2020; Madjid et al., 2020; Ronco and Reis, 2020). The severity of COVID-19 correlates with increased age, comorbidity factors such as diabetes, obesity, chronic lung disease, and cardiovascular disease, and male sex.
although severe disease is not limited to these risk groups (Guo et al., 2020; Chen et al., 2020b; Garg et al., 2020; Yang et al., 2020a).

4. Pathogenesis of COVID-19

4.1. SARS-CoV-2 infection

The epithelium in the respiratory tract functions as a physical barrier and also contributes to the elimination of invading pathogens. The alveoli in the lower respiratory tract are lined with type I alveolar epithelial cells (AEC I) which cover 90–95% of the alveolar surface and regulate gas exchange. In contrast, type II alveolar epithelial cells (AEC II) represent only about 7% of the alveolar surface and produce antimicrobial products such as complement, lysozyme, and surfactant proteins (SP). Although lung inflammation causes the most prominent symptoms of COVID-19, ACE2 mRNA expression in the lung is only moderate and concentrated in AEC II cells (Li et al., 2020; Zhao et al., 2020). Similar to SARS-CoV, the novel coronavirus may also nonproductively infect immune cells such as T cells (Gu et al., 2005), macrophages (Cheung et al., 2005; Yilla et al., 2005), monocytes (Yilla et al., 2005) and dendritic cells (DCs) (Law et al., 2005). SARS-CoV-2 may induce phenotypic and functional maturation of cells of the mononuclear phagocyte system (MPS) as SARS-CoV has been shown to upregulate MHC class II and costimulatory molecules on monocyte-derived DCs (Tseng et al., 2005). The MPS comprises not only DCs but also macrophages and monocytes. These immune cells are similar with regard to ontogeny, location, function and phenotype (Guilliams et al., 2014). Intriguingly, MPS cells account for the majority of immune cells in the inflamed lung microenvironment of severe COVID-19 (Liao et al., 2020). SARS-CoV-2 may use MPS cells as a Trojan horse in order to disseminate within the human organism and establish systemic infection (Park, 2020) similar to other emerging pathogens such as hantaviruses (Raftery et al., 2002; Raftery et al., 2020).

Although SARS-CoV-2 primarily replicates in the respiratory tract, autopsies demonstrate that it can infect cells in multiple organs, including the lungs, pharynx, heart, liver, brain, and kidneys (Puelles et al., 2020). Expression level and tissue distribution of ACE2 and TMPRSS2 mark the route of viral spread throughout the body (Sungnak et al., 2020). Besides lung epithelial cells other cells in the kidney, intestine, and heart express also ACE2 and can be infected (Zou et al., 2020; Hamming et al., 2004; Farkash et al., 2020; Hikmet et al., 2020; Zhao et al., 2020; Ziegler et al., 2020). Importantly, endothelial cells express ACE2 (Hamming et al., 2004; Ferrario et al., 2020; Hikmet et al., 2020) and are susceptible to infection with SARS-CoV-2 (Varga et al., 2020). In accordance, infection of human blood vessel organoids (Law et al., 2020a; Zhao et al., 2020) and MPS cells may account for the majority of immune cells in the inflamed lung microenvironment of severe COVID-19 (Liao et al., 2020). SARS-CoV-2 may use MPS cells as a Trojan horse in order to disseminate within the human organism and establish systemic infection (Park, 2020) similar to other emerging pathogens such as hantaviruses (Raftery et al., 2002; Raftery et al., 2020).

4.2. Megakaryocytes and platelets

It is becoming apparent that clotting at multiple sites including the brain plays an important role in COVID-19 pathogenesis (Willyard, 2020; Wise, 2020). In most of the terminally ill COVID-19 patients disseminated intravascular coagulation (DIC) is found (Tang et al., 2020) and may play a major role in the development of multi organ failure (Gando et al., 2020). In accordance, high blood levels of D-dimers, which are produced when protective mechanisms try to dissolve clots, can predict a severe course of COVID-19 (Zhang et al., 2020b). In fact, the extent of thrombosis is remarkably high affecting 20–40% of COVID-19 patients that are critical ill (Klok et al., 2020; Middeldorp et al., 2020; Poissy et al., 2020). The underlying mechanisms are unclear but most likely involve virus-induced inflammation of vessels (Wise, 2020). In addition, a drop in platelet numbers occurs and is associated with a severe course of COVID-19 and higher lethality (Lippi et al., 2020). Altered thrombopoiesis and increased destruction, consumption and sequestration of platelets may all contribute to thrombocytopenia in severe COVID-19 (Xu et al., 2020).

Platelet-releasing megakaryocytes reside not only in the bone marrow but also in the vasculature of the lung (Lefrancais et al., 2017). It is estimated that up to 60% of total platelet production in humans is due to platelet biogenesis in the lung (Lefrancais and Looney, 2019; Washington et al., 2020). Intriguingly, histopathologic analyses of SARS-CoV-2-related death revealed platelet-fibrin thrombosis and intravascular megakaryocytes in many organs, including the heart, lungs, kidneys, and liver (Fox et al., 2020). The pathogenic role of these broadly distributed extramedullary megakaryocytes in COVID-19 is unclear at the moment. It is likely, however, that the observed thrombosis in microvessels of COVID-19 patients is due to immunothrombosis. This form of thrombosis, which is triggered by inflammatory stimuli, aims at immobilizing invading pathogens and may represent an evolutionarily ancient component of the innate immune system in mammals (Engelmann and Massberg, 2013; Gaertner and Massberg, 2016). Thus, an inadequate innate immune response plays a crucial role in the pathogenesis of COVID-19.

4.3. Innate immune response

The innate immune response in the lung is a well ordered process in which several tiers of defense are involved (IWASAKI et al., 2017). At the site of infection in the respiratory tract, invading pathogens are detected by specialized cells such as airway epithelial cells (AEC), mast cells and MPS cells. These sensor cells are well equipped with pattern recognition receptors (PRRs) such as Toll-like receptors (TLRs), retinoic acid-inducible gene I (RIG-I) and other RIG-I-like receptors (RLRs) (Chow et al., 2018). Pathogen-associated molecular patterns (PAMPs) derived from viruses trigger a specific combination of PRRs and adaptor molecules resulting ideally in an immune response tailor-made for the pathogen. For example, double-stranded RNA (dsRNA) represents a prototypical virus-derived PAMP that is also generated during coronavirus replication and detected by RIG-I (Kindler et al., 2016). In addition, replication of cytopathic viruses such as coronaviruses cause multiple changes in cellular homeostasis such as
accumulation of misfolded protein aggregates, lysosomal disruption, mitochondria damage and imbalanced ion concentrations (Zhao and Zhao, 2020). This may trigger pyroptosis, a necrotic form of programmed cell death, which ushers into secretion of pro-inflammatory molecules of the IL-1 family (Chen et al., 2019; Tsuchiya, 2020). Virus-induced cell death also releases intracellular damage-associated molecular patterns (DAMPs) such as high-mobility group box 1 (HMGB1) and histones, which are normally hidden from recognition by PRRs (Schafer, 2014). However, upon release DAMPs trigger classical PRRs in neighboring epithelial and endothelial cells to generate additional pro-inflammatory cytokines and chemokines including IL-6, IP-10, macrophage inflammatory protein 1α (MIP1α), MIP1β and MCP1 (Tay et al., 2020). These then attract more effector cells (MPS cells and T cells) thereby driving the inflammatory process.

Sensing of coronaviruses by RIG-I and other PRRs starts an innate immune response that - if not blocked by the virus - would efficiently restrict viral replication (Tay et al., 2020). First-order cytokines such as IFN-α (IFN-α and IFN-β) and IFN-III are released and help to control and eliminate virus infections (Lazar et al., 2019). They may directly clear virus from infected cells by activating IFN-stimulated genes (ISGs) that exert direct antiviral activities or recruit antiviral effector immune cells such as macrophages. On the other hand, the antiviral immune response is a double-edged sword: It may be harmful for the organism if the timing, vigor, and target tissue of the immune response is not adequate as often observed during infection with zoonotic viruses (Supramaniam et al., 2018).

### 4.3.1. Interferons and proinflammatory cytokines

IFN-Ⅰ/IFN-Ⅲ are important antiviral cytokines, which induce a similar but not identical expression pattern of ISGs (Lazar et al., 2019). Due to different receptor usage, nearly every nucleated cell responds to type I IFN, whereas the response to type III IFNs is restricted to barrier tissues such as mucosal surfaces in the respiratory tract (Wack et al., 2015). A plethora of studies have reported an emerging role of IFN-Ⅲ in the first line defenses against respiratory viral infections (Andreakos et al., 2017; Galani et al., 2017) making IFN-Ⅲ a candidate for prevention and treatment of COVID-19 (Prokunina-Olsson et al., 2020). Supporting this notion, prophylactic and therapeutic treatment with IFN-Ⅲ in a mouse model of COVID-19 protected mice from the deleterious consequences of infection (Dinon et al., 2020). It has to be taken into account, however, that in mouse models IFN-Ⅲ can also have detrimental effects during long-term treatment and if considerable lung damage already exists before IFN-Ⅲ application (Broggi et al., 2020).

To study the innate response to SARS-CoV-2 near the site of infection, transcriptomic profiling of cells in bronchoalveolar lavages (BAL) from COVID-19 patients was performed (Zhou et al., 2020c). Bronchoscopic BAL is a useful technique for collecting lower respiratory tract specimens at the alveolar level from different areas of the lung and can predict the etiology of pneumonia in critical ill patients admitted to the ICU (Choi et al., 2014). A recent study reported elevated levels of both pro-inflammatory cytokines and IFN-Ⅰ/Ⅲ in BAL fluid from severe COVID-19 cases (Broggi et al., 2020). In contrast, Zhou et al. found no significant upregulation of IFN-Ⅰ in BAL cells although robust ISG induction was detected (Zhou et al., 2020c). In accordance, compared to other respiratory viruses, SARS-CoV-2 replication induces unusually low levels of interferon IFN-Ⅰ and IFN-Ⅲ both in cell culture and in ferrets, which serve as an animal model. A similar finding has been reported in post mortem lung samples from COVID-19 patients (Blanco-Melo et al., 2020). In ex vivo human lung tissue explants, SARS-CoV-2 did not significantly induce IFN-Ⅰ despite more efficient replication in comparison to SARS-CoV (Chu et al., 2020). Finally, an impaired IFN-Ⅰ response in association with a high virus load was observed in the blood of severe and critical COVID-19 patients (Hadjad et al., 2020).

These clinical and experimental observations strongly suggest that SARS-CoV-2 can efficiently subvert induction of IFN-Ⅰ/Ⅲ in infected cells as demonstrated for SARS-CoV (Chow et al., 2018). A recent report demonstrates that the nonstructural protein 1 (Nsp1) of SARS-CoV-2 interferes with RIG-I dependent innate immune responses that would otherwise facilitate production of IFN-Ⅰ/Ⅲ and virus elimination (Thoms et al., 2020). Furthermore, replication of genomic and subgenomic coronavirus RNA takes place in a special double membrane compartment that separates viral PAMPs from important PRRs such as RIG-I (Frieman and Baric, 2008). This compartmentalization also decreases detection of viral replication by cytoplasmic sensors. Finally, activation of NF-κB impairs IFN-Ⅰ signaling thereby facilitating viral replication (Wei et al., 2006; Pauli et al., 2008) suggesting that cross-regulation between IFN-Ⅰ/Ⅲ and NF-κB signaling cascades exists (Smits et al., 2010).

Low levels of IFN-Ⅰ/Ⅲ allow prolonged viral replication that in turn facilitates oxidative stress. The latter is often induced by respiratory viruses (Khomich et al., 2018) and oxidatively modified proteins are found in BAL derived from ARDS patients or patients at risk of ARDS (Lenz et al., 1999). It reflects an imbalance between generation of reactive oxygen species (ROS) by enzymes such as NADPH oxidases, and scavenging of ROS by endogenous antioxidants (Chatterjee, 2016). This imbalance can divert from virus-specific, IFN-Ⅰ/Ⅲ driven innate immune responses and result in activation of compensatory but less-specific antiviral responses driven by the redox-sensitive transcription factor NF-κB (Schreck et al., 1991). Moreover, high ROS levels trigger oxidation of proteins, lipids and DNA, which subsequently may act as DAMPs that foster inflammation and tissue injury (Imai et al., 2005; Shimada et al., 2012). Virus-induced oxidative stress in conjunction with necrosis of virus-infected cells leads to the generation and release of oxidized endogenous ligands that function as strong DAMPs and are sensed by TLRs (Gill et al., 2010). In a mouse model of virus-induced ALI, oxidative stress triggers lung injury by upregulating production of NF-κB driven proinflammatory cytokines such as TNF-α, IL-1β, or IL-8 and adhesion molecules (Imai et al., 2008). Diversion of antiviral innate immunity by oxidative stress can foster pathological inflammation and unleash a “cytokine storm”, in which MPS cells play a crucial role (Merad and Martin, 2020). Indeed, levels of NF-κB-driven proinflammatory cytokines such as TNF-α, IL-6, IL-8 (CXCL8), GM-CSF and G-CSF as well as chemokines such as MCP1, IP10 and MIP1α are also strongly enhanced in human individuals and animal models after infection with SARS-CoV-2 (Blanco-Melo et al., 2020; Chen et al., 2020a, 2020c; Hadjadj et al., 2020; Huang et al., 2020; Liu et al., 2020a; Qin et al., 2020; Yang et al., 2020b; Zhou et al., 2020c). In uninfected MPS cells, the NF-κB pathway may be triggered by interaction of the viral S-protein with receptor molecules on the cell surface (Dosch et al., 2009). High amounts of anti-inflammatory cytokines such as the IL-10 family cytokines are also detected in severe COVID-19 (Chen et al., 2020a) and are induced as part of a negative feedback loop (Ouyang and O’garra,
In mouse models of coronavirus-induced ALI, it has been shown that the timing of the IFN-I response relative to the kinetics of virus replication is critical in determining the outcome of infection with MERS-CoV (Channappanavar et al., 2019). Early IFN-I administration was protective. In contrast, late administration enhanced lethality by facilitating recruitment of neutrophils and inflammatory MPS cells, thereby increasing the release of proinflammatory cytokines, and impairing virus-specific T cell responses (Channappanavar et al., 2019). Similarly, in a mouse model of SARS-CoV infection the endogenous IFN-I response was delayed and lagged behind peak virus replication resulting in a deleterious effect of IFN-I (Channappanavar et al., 2016). Supporting a crucial role for NF-κB-driven inflammation, inhibition of NF-κB signaling in mice infected with SARS-CoV reduced inflammatory lung pathology and significantly enhanced survival (Dediego et al., 2014).

In aged experimental animals, SARS-CoV triggers a much stronger NF-κB driven innate response than in younger animals resulting in exacerbated ALI despite similar levels of viral replication (Baas et al., 2008; Rockx et al., 2009; Smits et al., 2010). This could explain why the severity of COVID-19 increases with age (Chen et al., 2020b) if SARS-CoV-2 does the same in humans. This effect is best explained by an age-related increase in ROS levels that reflect a compensatory homeostatic response to maintain physiological intracellular signaling pathways during aging (Lopez-Otin et al., 2013). It is well described that ROS have not only tissue damaging effects but also play an important physiological role in regulating various cellular processes including differentiation, autophagy, metabolic adaptation, and immune cell activation (Sena and Chandel, 2012). During infection with pathogenic respiratory viruses such as SARS-CoV-2, however, ROS levels in elder people may further increase and reach a certain threshold resulting in overactivation of NF-κB and inflammatory tissue damage (Chung et al., 2006). Similarly, the higher susceptibility of men to oxidative stress (Kander et al., 2017) may contribute to the fact that men are more prone to severe COVID-19 than women (Grasselli et al., 2020; Richardson et al., 2020).

Altogether, it is likely that the delayed and insufficient IFN-I response to SARS-CoV-2 allows prolonged virus replication and increased oxidative stress, which triggers a NF-κB-driven cytokine storm resulting in excessive inflammation.

4.3.2. Neutrophils and neutrophil extracellular traps (NETs)

Intriguingly, transcriptome analysis of human lung epithelial cells infected with SARS-CoV-2 in vitro and analysis of cytokines/chemokines in sera from COVID-19 patients revealed overexpression of several NF-κB-driven chemokines that attract neutrophils, e.g. CXCL2 and CXCL8 (Blanco-Melo et al., 2020). Increased levels of neutrophil attractant proteins such as CXCL8 are also detected in patients severely infected with SARS-CoV (Wong et al., 2004), MERS-CoV (Min et al., 2016), or SARS-CoV-2 (Chen et al., 2020a; Hadjadj et al., 2020; Huang et al., 2020; Liu et al., 2020; Qin et al., 2020; Yang et al., 2020; Zhou et al., 2020c). Neutrophils are the most abundant immune cells and represent about 50–70% of all circulating leukocytes (Ley et al., 2018). Under physiological conditions approximately 10¹¹ neutrophils are generated each day in the human organism (Dancey et al., 1976). They are short-lived and act as first line of defense against invading pathogens (Bardocz et al., 2014; Nemeth et al., 2020). A body of evidence now exists showing that neutrophils also play an important role in combating viral infections (Naumenko et al., 2018). Large numbers of neutrophils have been shown to accumulate within the pulmonary microcirculation in the steady state suggesting that respiratory viruses interact with this important cell type at a very early stage of infection (Devi et al., 2013; Casanova-Acebes et al., 2018). Finally, activated neutrophils are of central importance to the pathogenesis of ALI, e.g. by releasing proteolytic enzymes or ROS (Lee and Downey, 2001; Grommes and Soehnlein, 2011).

Patients with severe COVID-19 are often lymphopenic whereas the number of neutrophils is increased (Fox et al., 2020; Wang et al., 2020a). There is ample evidence that an increased neutrophil-to-lymphocyte ratio is an independent risk factor precipitating a severe course of COVID-19 (Liu et al., 2020; Qin et al., 2020; Song et al., 2020; Zhang et al., 2020a). Neutrophils and platelets closely interact to fulfill their functions in innate immune responses (Kim and Jenne, 2016). The formation of neutrophil-platelet complexes facilitates recruitment of neutrophils to inflamed tissue (Sreeramkumar et al., 2014). Moreover, neutrophils show a more vigorous defense against pathogens and release higher amounts of neutrophil mediators after binding to platelets. Finally, platelets can induce neutrophils to produce neutrophil extracellular traps (NETs) that protect from viral infection (Jenne et al., 2013). NETs are formed when neutrophils undergo a form of programmed cell death (Takei et al., 1996; Brinkmann et al., 2004) that has also been referred to as NETosis. However, the demarcation of NETosis from other forms of cell death is a matter of debate (Boelitz et al., 2019). Thus, in this review the term “NET formation” is used in preference to NETosis. NET formation may also occur without disruption of cellular membranes preserving neutrophil functions such as chemotaxis and phagocytosis (Clark et al., 2007; Yipp et al., 2012). NETs are net-like structures composed of chromatin festooned with neutrophil granule proteins. Several enzymes are involved in the process of NET formation including neutrophil elastase (NE), which enters the nucleus, degrades nuclear proteins and initiates nuclear disintegration (Papayannopoulos, 2018).

Analysis of autopsy samples from the lungs of COVID-19 patients revealed neutrophil infiltration in pulmonary capillaries, acute capillaritis with fibrin deposition, extravasation of neutrophils into the alveolar space, and neutrophilic mucositis (Barnes et al., 2020). A prominent feature of COVID-19 is multiorgan inflammation with vessel walls containing neutrophils (Varga et al., 2020). Within alveolar capillaries of patients with COVID-19, neutrophils and platelets get entrapped in fibrin meshworks (Fox et al., 2020). These neutrophils are partially degenerated suggesting that NETs are generated (Fox et al., 2020). Indeed, high serum levels of cell-free DNA, DNA-myeloperoxidase (MPO) complexes, and citrullinated histone H3 (Cit-H3) have been detected in COVID-19 and correlate with disease severity (Zuo et al., 2020a). Viable SARS-CoV-2 can directly induce NET formation in healthy neutrophils by an unknown mechanism (Veras et al., 2020). MPO-DNA and Cit-H3 are regarded as specific markers for NETs suggesting that NETs may drive cytokine release, respiratory failure, and microvascular injury in COVID-19. Several viruses including respiratory viruses reportedly induce NETs, which subsequently bind, entrap and neutralize virions, while contributing to immune activation via pattern recognition
response of the adaptive immune system only occurs within several days. An efficient long lasting and antigen specific immune defense by the adaptive immune system for novel therapeutic strategies.

4.4. Antigen specific immune defense by the adaptive immune system

While the activation of the innate immune system as a first line of defense starts within minutes to hours, an effective immune response of the adaptive immune system only occurs within several days. An efficient long lasting and antigen specific immune response of the adaptive immune system depends on the generation of antigen specific T-helper cells that activate B-cells to produce specific antibodies. The antibodies then mediate the destruction of the pathogen through complement activation or antibody dependent cellular cytotoxicity.

In addition, the adaptive immune system activates antigen specific cytotoxic T-cells that specifically kill infected cells. This process requires the presentation of antigen by antigen presenting cells such as dendritic cells and macrophages to T-cells. The T-cells then differentiate into either helper T-cells that stimulate other immune cells, or cytotoxic T-cells that directly kill infected cells.

The adaptive immune response is specific to the type of pathogen and provides long lasting protection against future infections. This is achieved through the generation of memory B-cells and T-cells that can rapidly respond to subsequent exposures to the same pathogen. This memory response is crucial for the prevention of severe infections that can be life threatening.

The development of antigen specific T-helper cells occurs in the lymph nodes and proceeds through several stages. Initially, antigen specific T-helper cells are generated in the thymus, a specialized organ in the lymphoid system, where T-cells develop from hematopoietic stem cells. These T-cells then enter the lymph nodes where they encounter specific antigens presented by antigen presenting cells. The encounter between the T-cell and the antigen is determined by the interaction of the T-cell receptor with the major histocompatibility complex (MHC) that presents the antigen.

The T-cell receptor signals the T-cell and activates processes that lead to the differentiation of the T-cell into a specific helper cell. These helper cells then interact with B-cells, which are activated to produce antibodies specific to the pathogen. The antibodies then mediate the destruction of the pathogen through complement activation or antibody dependent cellular cytotoxicity.

The development of antigen specific cytotoxic T-cells similarly occurs in the lymph nodes where T-cells interact with antigen presenting cells. The interaction between the T-cell receptor and the MHC antigen complex triggers a signaling pathway that leads to the differentiation of the T-cell into a specific cytotoxic cell. These cytotoxic cells then directly kill infected cells.

The adaptive immune response is crucial for the protection against infections and the prevention of severe infections that can be life threatening. The development of antigen specific T-cells and B-cells is an important target for the development of vaccines that can induce a strong immune response against specific pathogens.
reaction, however, is essential to control viral infections in the long term and to prevent virus persistence. In viral infections, effector CD8\(^+\) T cells play an important role in killing virus-infected cells, whereas CD4\(^+\) T cells provide help to effector CD8\(^+\) T cells and B cells. The latter produce neutralizing antibodies that are important for viral clearance and long-lasting antiviral immunity. Both CD4\(^+\) and CD8\(^+\) T cells produce IFN-γ, which exerts its antiviral effects not only by inducing ISGs but also by regulating the antiviral immune response (Boehm et al., 1997). A third type of T cells, the regulatory T cells (T\(_{reg}\)), counteracts these activities when necessary, for example to limit the time of activation (Li et al., 2018). Mostly, T\(_{reg}\) belong to the CD4\(^+\) T cell population and express FoxP3 (forkhead box P3), however, some of them are also CD8\(^+\) T cells. Expression of the anti-inflammatory cytokine IL-10 enhances the activity of T\(_{reg}\) cells leading to immunosuppression and T cell anergy. T\(_{reg}\) also counteract inflammatory processes that are induced by cells of the innate immune system. They inhibit IL-6 production of neutrophils and dictate that neutrophils produce anti-inflammatory molecules (IL-10, TGF-β and indoleamine 2,3-dioxygenase (IDO)) (Lewkowicz et al., 2013). Through the release of TGF-β, T\(_{reg}\) induce neutrophil apoptosis. In this way, they can contribute to the resolution of an acute lung injury (D’alessio et al., 2009). T\(_{reg}\) can also inhibit neutrophil infiltration into injured tissue. This improved the healing of myocardial infarction and kidney ischemia-reperfusion injury (Kinsey et al., 2009; Weirather et al., 2014). It can therefore be assumed that T\(_{reg}\) also play an important role in tissue regeneration in COVID-19.

**Fig. 1. Basic features of the immunopathogenesis in COVID-19 with central role of ROS:** Both innate (right site) and adaptive immune responses (left site) are involved in immunopathogenesis of COVID-19. The pathogenic cascade starts with prolonged and extensive replication of SARS-CoV-2 in lung epithelial cells and in endothelial cells of the vessels due to viral evasion of the IFN-I/III response. As a consequence, neutrophils and MPS cells (e.g. macrophages, monocytes, and immature DCs) are massively recruited to the inflammatory tissue. Activated neutrophils and MPS cells produce large amounts of ROS thereby creating an imbalanced oxidative stress response. Oxidation of endogenous molecules (e.g. DNA and lipids) results in DAMPs that trigger pro-inflammatory cytokine secretion (cytokine storm) through TLR signaling thereby activating the redox-sensitive transcription factor NF-κB. ROS and TLR signaling also induce an overflow of NETs. There may be several positive feedback loops between cytokines (TNF-α, IL-1β) and ROS production as well as between cytokines (TNF-α, IL-1β) and NET formation. ROS, NETs and proteolytic enzymes released by activated neutrophils also contribute to organ damage and clotting in vessels. On the other side, imbalanced ROS production also suppresses the T cell response thereby contributing to lymphopenia in COVID-19. As a result, less activated antiviral CD8 T cells are available to kill virus-infected cells and to eliminate the virus. Moreover, CD4\(^+\) T cells are less efficient in helping B cells to produce neutralizing antibodies and establish long-term immunity.
Sustained stimulation by viral antigens can lead to exhaustion of the T cells, which are responsible for eliminating the viruses (Schönrich and Raftery, 2019). Exhausted T cells (T_{ex}) express programmed cell death protein 1 (PD-1) and/or T cell immunoglobulin and mucin domain-3 (TIM-3) and are finally depleted by apoptosis. Blockade of IL-10 or checkpoint inhibitors that block the PD-1 pathway can at least in part revert T cell exhaustion.

### 4.4.1. Lymphopenia and failure of antiviral T cell responses in severe COVID-19

Systemic viral infections are usually accompanied by lymphocytosis, an increase in lymphocytes in the blood. This is due to the fact that the pool of antigen specific CD8\(^+\) T lymphocytes needs to be enlarged. In COVID-19, however, decreased numbers of CD4\(^+\) and CD8\(^+\) T cells and low IFN-γ production by CD4\(^+\) T cells are observed (Chen et al., 2020a; Liu et al., 2020a; Wang et al., 2020a). The numbers of CD8\(^+\) T cells appear as independent predictor for COVID-19 severity and treatment efficacy (Wang et al., 2020b). In fact, the subacute progression of COVID-19 implicates that both T cell depletion and T cell exhaustion contribute to the failure of the adaptive immune system (Vardhana and Wolchok, 2020). In accordance, T cells from COVID-19 patients show strongly upregulated PD-1 and TIM-3 (Diao et al., 2020). The resulting weak adaptive immune response permits sustained virus replication, which further drives immunopathogenesis through the excessive inflammatory activity of the innate immune cells, a vicious cycle. In line with this assumption, the severity of the disease correlates with an increase in the neutrophil to lymphocyte ratio (Liu et al., 2020b; Qin et al., 2020; Song et al., 2020; Zhang et al., 2020a). At the moment, however, the precise mechanisms contributing to the failure of the antiviral T cell response in COVID-19 are enigmatic.

### 4.4.2. Oxidative stress as potential cause of lymphopenia and T cell suppression in COVID-19

One potential mechanism for COVID-19-associated reduction in T cell numbers is apoptosis. Chu et al. showed that MERS-CoV, but not SARS-CoV, can efficiently infect human T cells and trigger their apoptosis via both extrinsic and intrinsic apoptosis pathways (Chu et al., 2016). SARS-CoV-2 can induce lymphocyte apoptosis via enhancing Fas signaling (Chen et al., 2020d). In addition to Fas-induced apoptosis by the virus, a reduction in T cell numbers can also be due to oxidative stress (Klemke et al., 2008; Wabnitz et al., 2010), which is typically present in COVID-19 and ARDS (Imai et al., 2008; Kellner et al., 2017). A pro-oxidative milieu leads to oxidation of essential regulatory proteins in T cells, such as cofilin (Klemke et al., 2008; Wabnitz et al., 2010; Samstag et al., 2013) or L-plastin (Balta et al., 2019). Consequently, T cells become hypo-responsive or even die. As described above, activated neutrophils and MPS cells are to a large extent responsible for the massive release of ROS into the lung tissue. In addition, the massive TNF-α release during the cytokine storm described above could exacerbate ROS production via a positive feedback loop by activating NADPH oxidases (Blaser et al., 2016). TNF-α induced ROS production could also contribute to the extension of COVID-19 symptoms to distant tissues such as the brain (Sándoval et al., 2018).

It is estimated that more than 1.7 billion people (more than 20% of the world’s population) have a health problem that increases the risk for severe COVID-19 such as cardiovascular disease or diabetes mellitus (Clark et al., 2020). In the pathogenesis of these comorbidities imbalanced ROS production is also key (Shah and Brownlee, 2016), supporting the importance of oxidative stress for severe COVID-19. High glucose and also hypoxia/reperfusion occurring upon ventilation of COVID-19 patients promote ROS production. This can induce NLRP3 inflammasome-mediated pyroptosis, which is further enhanced in the presence of the bacterial component LPS (Shah and Brownlee, 2016). Moreover, SARS-CoV papain like protein significantly triggered a ROS/p38 MAPK/STAT3 pathway, leading to activation of the TGF-β1 promoter in lung epithelial cells. This effect correlated with up-regulation of pro-fibrotic responses in vitro and in vivo (Li et al., 2016). A ROS mediated release of TGF-β could also contribute to the observed lymphopenia in COVID-19, since TGF-β is a potent immunosuppressive substance acting on T cells. Finally, a pro-oxidative state in T cells (increase in ROS and decrease in GSH) was shown to promote the development of T_{reg} cells (Liang et al., 2020). A relative increase in T_{reg} would also counteract the T cell defense of SARS-CoV-2. Consistent with this assumption, it was reported that, unlike other T cells, the T_{reg} population in COVID-19 patients was not decreased (Shi et al., 2020b). Overall, oxidative stress likely plays a central role in the pathogenesis of severe COVID-19 (Fig. 1).

### 5. Treatment options for COVID-19 associated immunopathogenesis

The precise understanding of COVID-19 associated immunopathogenesis as summarized in Fig. 1 paves the way for developing novel treatment options. Approaches that interfere with neutrophil recruitment, neutrophil activation, and NET formation have been discussed in detail elsewhere (Barnes et al., 2020; Narasaraju et al., 2020). In addition, ROS production, cytokine storm, and T cell suppression can all be targeted specifically to ameliorate the course of SARS-CoV-2 infection. For reasons of space, we will concentrate in this overview on the antagonization of ROS and proinflammatory cytokines, which represent key targets to interrupt the vicious cycle that drives severe COVID-19.

#### 5.1. Antioxidative substances could improve the survival and function of T cells in COVID-19 patients and prevent NET formation

##### 5.1.1. Vitamin C

Recently, it has been hypothesized that vitamin C, also known as ascorbic acid, could have a protective effect that prevents ALI in sepsis (Li, 2018). In mouse models, vitamin C infusion indeed protected from the deleterious consequences of sepsis (Fisher et al., 2011). It enhanced the epithelial barrier function, increased alveolar fluid clearance, attenuated the proinflammatory response, and prevented sepsis-associated coagulation abnormalities (Fisher et al., 2012). Importantly, vitamin C prevented the activation of NF-kB and neutrophil sequestration (Fisher et al., 2011). In line with these findings, antioxidant vitamins including vitamin C protected...
against ARDS in a rat model (Erol et al., 2019). A clinical trial in sepsis patients showed that a high dose of vitamin C infusion is well tolerated in sepsis patients (Fowler et al., 2014). However, vitamin C infusion in a randomized study in patients with sepsis and ARDS did not improve organ dysfunction or the markers for inflammation and vascular injury (Fowler et al., 2019). Yet, in contrast to the mouse experiments described above, the vitamin C infusion in the patients was only carried out at a very late stage of the disease, in which the harmful effects of oxidative stress had probably already occurred.

For COVID-19, treatment with vitamin C still appears to be an attractive option. In this context, it is interesting to note that patients with infections often have a reduced vitamin C level due to metabolic changes (Hemilä, 2017). Vitamin C has multiple effects on the development, proliferation and function of lymphocytes. It has been shown to promote differentiation and proliferation of T cells (Manning et al., 2013; Ang et al., 2018; Van Gorkom et al., 2018; Kouakanou et al., 2020). When melanoma primed DCs were pre-incubated with vitamin C before co-culture with T cells, an enhanced generation of melanoma-specific effector and effector memory CD8+ T cells was found. This was accompanied by an increased killing of melanoma cells (Jeong et al., 2014). Such an effect would be helpful in COVID-19, since the decrease in CD8+ T cells was a predictor for COVID-19 severity and treatment efficacy (Wang et al., 2020b). Besides acting as an antioxidant, parts of the effects of vitamin C were due to its influence on epigenetic regulation (e.g. enhancement of histone demethylation). In addition, it inhibited the activation of NLRP3 inflammasomes in a macrophage cell line by blocking the intracellular shuttling of thioredoxin-interacting protein (TXNIP), a key molecule of cellular redox regulation (Choe and Kim, 2017). It was also found that the presence of vitamin C is necessary for the apoptosis and clearance of dead neutrophils from infection sites by macrophages (Carr and Maggini, 2017). Furthermore, vitamin C significantly weakened NET formation in healthy people (Bozozet and Carr, 2019). Infusion of vitamin C, therefore, appears to be a promising option to counteract the oxidative stress that leads to NF-kB driven cytokine storm and NET formation and promotes ALI/ARDS.

In three controlled studies vitamin C reduced the incidence of pneumonia in common cold and an improvement of the pneumonia by the vitamin C treatment was reported in two controlled studies (Hemila and Louhiala, 2013). A prospective randomized placebo controlled triple masked clinical trial (https://clinicaltrials.gov/ct2/show/NCT04264533) launched in February 2020 in Wuhan, China, will show how vitamin C infusion affects severe COVID-19 (Carr, 2020). Additional clinical trials are on the way, e.g. in the USA (https://clinicaltrials.gov/ct2/show/NCT04344184) and in Italy (https://clinicaltrials.gov/ct2/show/NCT04323514). Contrary to what is planned in these clinical studies, vitamin C treatment should, however, be started earlier (between days 1 and 7, and especially before an intensive care unit is required), in order to prevent ROS-induced suppression of antiviral T cells and maintain a normal neutrophil to lymphocyte ratio. This may prevent SARS-CoV-2 infection from spreading and worsening towards ARDS.

5.1.2. N-acetylcysteine (NAC)

Likewise, another antioxidant may be an attractive treatment option for COVID-19, namely N-acetylcysteine (NAC). It is a well-known drug used for mucolysis and treatment of bronchitis and also inhibits NET formation by human neutrophils in vitro (Kirchner et al., 2013). Importantly, NAC was shown to prevent immunosuppression of T cells in a pro-oxidative environment (Liang et al., 2018) and may, therefore, be able to reverse lymphopenia in COVID-19.

NAC is a precursor of glutathione (GSH), a very important cellular antioxidant. Infection of differentiated normal human bronchial epithelial cell (NHBE) cultures with respiratory syncytial virus (RSV) significantly decreased the intracellular levels of GSH. Supplementation with NAC increased GSH and inhibited RSV infection, thereby restoring normal ciliary activity and inhibiting mucin release (Mata et al., 2012). NAC has also been tested in IAV infection suggesting that it attenuates symptoms and reduces the incidence of clinically apparent disease (De Flora et al., 1997; Lai et al., 2010). In a randomized controlled trial (RCT) with 39 patients in China suffering from community-acquired pneumonia (CAP), which is usually caused by bacteria or viruses, high dose NAC improved oxidative stress parameters (reduced malondialdehyde (MDA) and increased total antioxidant capacity (TAOC)) and diminished inflammatory factors (TNF-α). NAC-related adverse effects did not occur. Yet, the CT score of pneumonia did not change (Zhang et al., 2018). A phase II clinical trial has started at the beginning of May to treat severe or critically ill COVID-19 patients with NAC. This will be a non-randomized and non-masked single-institution study at Memorial Sloan Kettering Cancer Center, USA (https://clinicaltrials.gov/ct2/show/NCT04374461). An additional study that is planned by the Cambridge Health Alliance, USA, aims to determine whether NAC treatment can prevent those with mild or moderate COVID-19 from progressing to severe disease (https://clinicaltrials.gov/ct2/show/record/NCT04419025).

5.2. Blockade of proinflammatory cytokines

Finally, one further option to prevent imbalanced ROS production and a pathological cytokine storm of COVID-19 is by blocking the effect of TNF-α (Feldmann et al., 2020) or other proinflammatory cytokines such as IL-6. The respective biologicals such as the TNF-α antibodies infliximab and adalimumab have a good safety profile and are already frequently used to treat chronic inflammatory diseases (Silva et al., 2010) without impairing clearance of respiratory viruses (Schett et al., 2020). In RSV infected mice, anti-TNF-α treatment actually diminished the perivascular and peribronchial infiltrates and reduced the destruction of the lung tissue (Hussell et al., 2001). It does not seem unlikely that the inhibition of TNF mediated ROS induction contributed to this beneficial effect, besides blocking other known TNF functions. Therefore, anti-TNF treatment should probably be started early in the course of COVID-19 to interrupt the pathogenesis.

The RECOVERY trial (https://clinicaltrials.gov/ct2/show/NCT04381936) recently reported as a major breakthrough that dexamethasone (DXM) saves lives of severe COVID-19 patients (Leford, 2020). DXM has a broad range of inhibitory effects on the immune system including inhibition of proinflammatory cytokine production (Tobler et al., 1992) by shifting TLR4 signaling away from NF-kB driven pro-inflammatory responses (Mogensen et al., 2008; Broering et al., 2011; Chuang et al., 2017; Curtale et al., 2017). This
mechanism could be important in COVID-19 as TLR4 has been identified as a key pathway in a mouse model of virus-induced ALI/ARDS (Imai et al., 2008). In accordance, DXM impairs NET formation induced by TLR4 signaling in vitro (Wan et al., 2017), downregulates NET levels in an animal model of asthma (Vargas et al., 2017), and is associated with reduced circulating NETs in patients with asthma (Gál et al., 2020). However, the precise mechanism underlying the beneficial effects of dexamethasone in severe COVID-19 has to be elucidated in future studies.

6. Concluding remarks

A severe course of COVID-19 is a result of both the direct cytolytic effects of SARS-CoV-2 and the deleterious consequences of the immune response. The latter comprises a virus-induced cytokine storm as well as imbalanced ROS production by cells of the MPS and neutrophils resulting in extensive NET formation, inflammation and tissue destruction. Moreover, excess ROS contribute to a suppression of the adaptive immune system due to paralysis or elimination of CD8+ cytotoxic T cells and CD4+ helper T cells. As a result, the killing of virus infected cells and an efficient production of antiviral antibodies needed for the development of a long-lasting antiviral immunity is disturbed. Given the central role of oxidative stress and formation of pro-inflammatory NETs for the pathogenesis of severe COVID-19, treatment strategies should aim at decreasing the net ROS levels and NET formation, e.g. by antioxidants such as vitamin C or NAC and/or by inhibiting ROS production, e.g. by antagonizing the activation of cells of the MPS and neutrophil granulocytes and/or by inhibiting TNF-α. This oxidative stress-based concept of COVID-19 pathogenesis and treatment should be validated in randomized controlled clinical studies. As preclinical animal models for SARS-CoV-2-associated disease mice can be used that express human ACE2 as a transgene (Bao et al., 2020) or after adenovirus-mediated transfer to lung cells (Hassan et al., 2020; Sun et al., 2020). In the future, it will be important to carry out more research into the influence of SARS-CoV-2 on various human immune cell populations in order to gain a deeper insight into the cellular and molecular mechanisms that promote the progression from mild to severe COVID-19. The detailed elucidation of immunopathogenic mechanisms induced by SARS-CoV-2 will pave the way for novel treatment options for COVID-19.

Declaration of competing interest

The authors declare no financial or commercial conflict of interest.

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References


Gando, S., Fujishima, S., Saitoh, D., Shiraishi, A., Yamakawa, K., Kushimoto, S., Ogura, H., Abe, T., Mayumi, T., Sasaki, J., Kotani, J., Takeyama, N., Tsuruta, R., 


Drosten, C., Gunther, S., Preiser, W., Van Der Werf, S., Brodt, H.R., Becker, S., Rabenau, H., Panning, M., Kolesnikova, L., Fouchier, R.A., Berger, A., Burguiere, A.M., 


Cinatl, J., Eickmann, M., Grywna, K., Kramme, S., Manuguerra, J.C., Muller, S., Rickerts, V., Sturmer, M., Vieth, S., Klenk, H.D., Osterhaus, A.D., 


Drosten, C., Gunther, S., Preiser, W., Van Der Werf, S., Brodt, H.R., Becker, S., Rabenau, H., Panning, M., Kolesnikova, L., Fouchier, R.A., Berger, A., Burguiere, A.M., 


Dallinga, B., van den Berg, D., Schouten, L., Groothuis, T., van Kleef, E., Govaert, B., de Vries, R., van der Zalm, J., de Vries, P., Schermerhorn, G., van den Bosch, H., 


Drosten, C., Gunther, S., Preiser, W., Van Der Werf, S., Brodt, H.R., Becker, S., Rabenau, H., Panning, M., Kolesnikova, L., Fouchier, R.A., Berger, A., Burguiere, A.M., 


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